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## **Photocatalytic activity of titanium dioxide against bacteria and viruses**

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## **Introduction**

Increasing concern about pandemics of disease-causing bacteria and viruses (e.g., swine influenza H1N1 and severe acute respiratory syndrome coronavirus (SARS-CoV)) and overall airborne pollution have attracted worldwide attention and spurred the development of air purification technologies [1,2]. Environmental contamination is a complex and intriguing problem involving the presence of contaminants in the form of particles (i.e., dust and smoke), biological agents, such as bacteria, molds and viruses, and other gaseous contaminants such as CO, CO<sub>2</sub>, NO<sub>x</sub>, SO<sub>x</sub> and volatile organic compounds (VOCs) [3].

Purification from contaminants can be achieved by means of disinfection or sterilization processes. In more detail, disinfection describes a process that eliminates many or all potentially pathogenic microorganisms, except bacterial spores, on inanimate objects. Instead, sterilization describes a process that destroys or eliminates all forms of microbial life and is carried out in health-care facilities by physical or chemical methods. Widely used air purification techniques for disinfection include the treatment with non-thermal plasma (NTP), thermal treatment, use of antimicrobial material-embedded filters, ultraviolet (UV) light, and photocatalysts [2–4]. In the NTP air-cleaning systems, energetic electrons excite, dissociate, and then ionize gas molecules, giving rise to chemically reactive species such as atomic oxygen, hydroxyl radicals, and ozone [4]. Such active species inactivate biotic and abiotic particles, eventually forming secondary pollutants (e.g., ozone, CO, or NO<sub>x</sub>) [5]. In thermal treatment, exposure to high temperature induces protein denaturation through disrupting the polypeptide structures, thus resulting in damage to microorganisms [6]. However, it may consume much power to apply thermal energy at high temperature [7]. Filtration systems, in which airborne biological particles are collected on the surface of a filter, are great options to overcome the limitations of the aforementioned techniques.

However, these antimicrobial material-embedded filters are generally effective in the short-term because of the accumulation of dust that progressively clogged them and cause a large pressure drop, so that they must be replaced regularly to prevent the possible re-introduction of airborne microorganisms into the environment all of a sudden [5].

Another approach to prevent the transmission of airborne-mediated disease relies on the inactivation of airborne pathogens by means UV light<sup>1</sup> [8,9]. This is a cost-effective and environmentally-compatible alternative to frequently used chemical processes. UV irradiation has been found to consume little energy as compared to thermal treatments and can be simply applied by installing and turning on a UV lamp. In this regard, UV lamps are often installed in the ceilings of

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<sup>1</sup> UV-A ( $\lambda = 315 - 400$  nm); UV-B ( $\lambda = 280 - 315$  nm) UV-C ( $\lambda = 100 - 280$  nm)

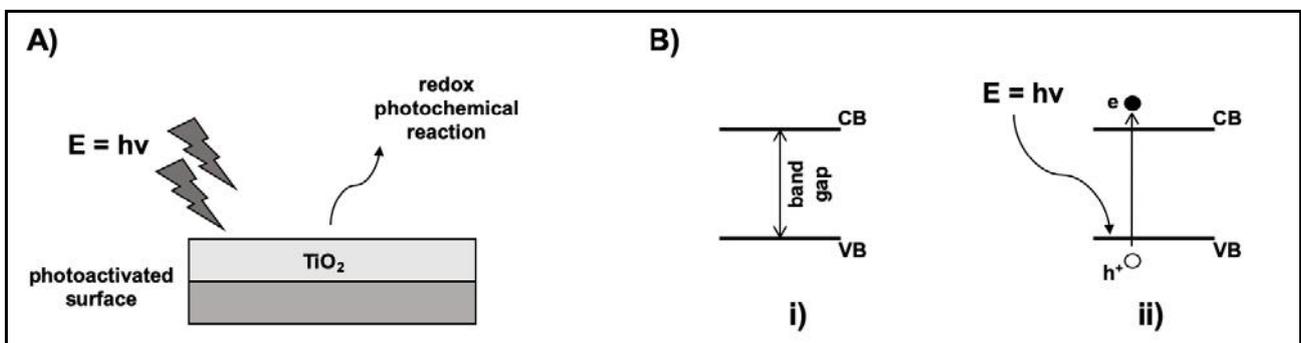


surgery rooms in hospitals and health care facilities and function to inactivate nearby bioaerosols [10]. Of note, germicidal UV light allows the inactivation of both drug-sensitive and multi-drug-resistant bacteria [11], as well as different viral strains [12]. However, the widespread use of germicidal UV lighting systems in public space has been very limited because conventional UV light sources are a human health hazard as well [13].

To date, one of the most promising technologies for environmental disinfection is photocatalysis (or UV photocatalytic oxidation (PCO)), which is one of the most important advanced oxidation technologies available [14]. PCO has many advantages over the other technologies, including the simultaneous treatment of mixtures of diverse pollutants, relatively low costs, and ease of operation and maintenance [1,15,16].

### 1. UV-activated photocatalysis: working principle

Photocatalysis using semiconductor (SC) has been proven to effectively degrade a vast array of pollutants. Although the detailed mechanism of photocatalysis varies with different pollutants, it is commonly agreed that the primary reactions responsible for the photocatalytic effect are interfacial red-ox reactions of electrons and holes that are generated when the SC catalyst is exposed to light of sufficient energy (**Figure 1A**). It is well known that a SC is characterized by a band energetic structure, with a band gap between the lower valence band (VB), entirely filled with electrons, and the unoccupied, higher energetic conduction band (CB). The adsorption of a photon with sufficient energy ( $h\nu$ ) by the SC promotes electrons from the valence band ( $e_{vb}^-$ ) to the conduction band ( $e_{cb}^-$ ), leaving a positively charged hole in the valence band ( $h_{vb}^+$ ) (**Figure 1B**). The electrons ( $e^\bullet$ ) are then free to migrate within the conduction band. The holes ( $h^+$ ) may be filled by the migration of an electron from an adjacent molecule, leaving the latter with a hole, and the process may be repeated.

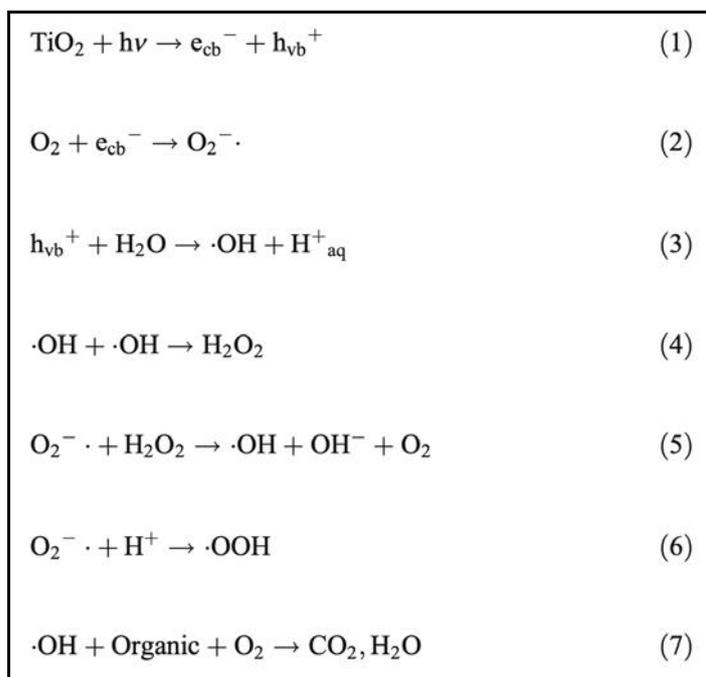


**Figure 1. A)** Schematic representation of a photoactivated antimicrobial surface. **B)** Band energetic structure (i) and mechanism of photoirradiation of a SC (ii).



Electrons and holes may recombine (bulk recombination) a non-productive reaction, or, when reaching the surface, they react to give reactive oxygen species (ROS) such as  $O_2^{\cdot-}$  (**Figure 2**, reaction 2) and  $\cdot OH$  (**Figure 2**, reaction 3). When in aqueous solution, these can react to give  $H_2O_2$  (**Figure 2**, reaction 4), further hydroxyl (**Figure 2**, reaction 5) and hydroperoxyl (**Figure 2**, reaction 6) radicals. Reaction of the radicals with organic compounds ultimately results in their mineralization (**Figure 2**, reaction 7), that is the degradation of organic contaminants due to their total oxidation [17,18].

Compared to other SC photocatalysts, titanium dioxide ( $TiO_2$ ) is the most promising material because of its high photoactivity behavior and stability, relatively low cost, and non-toxicity. For these reasons,  $TiO_2$  mediated photocatalytic inactivation of a large number of organic contaminants, including bacteria and viruses, has been extensively investigated [19–25]. Noteworthy, there are three main polymorphs of  $TiO_2$ : anatase, rutile and brookite. The majority of studies showed that anatase was the most effective photocatalyst and that rutile was the less active form. This is probably due to differences in the extent of recombination of electron and hole between the two forms. However, studies have shown that mixtures of anatase and rutile were more effective photocatalysts than 100% anatase [26]. In this context, Degussa P25 (Degussa Ltd., Germany) is a widely used, commercially available preparation of  $TiO_2$  which contains approx. 80% anatase and 20% rutile.



**Figure 2.** Mechanism of photocatalytic oxidation (POC). POC is based on the interaction between light and semiconductor particles, which produce the highly reactive oxygen species (ROS), such as  $OH^\bullet$ ,  $OH^-$ ,  $O_2^{\cdot-}$ ,  $HO_2^\bullet$ , capable of destroying organic (chemical and biological) contaminants



TiO<sub>2</sub> photocatalysts generate strong oxidizing power when illuminated with UV light with wavelengths ( $\lambda$ ) of less than 385 nm [27]. On the other hand, doping TiO<sub>2</sub> with C, N, S, and metals such as Sn, Pd, and Cu has been found to increase the  $\lambda$  radiation adsorption so that also visible light can be used to trigger the photocatalysis [28]. Generally speaking, photocatalytic TiO<sub>2</sub> particles can be used in the form of *i*) powder, usually dispersed in aqueous solutions, *ii*) film/coating applied to various substrates or *iii*) immobilized on surfaces [29–34].

As photocatalytic inactivation of contaminants is a synergistic bactericidal effect of UV light and oxidative radicals generated at the TiO<sub>2</sub>-based illuminated surface, some parameters, such as light intensity, extent of irradiation, catalyst concentration, play a role on the disinfection behavior. The UV dose (referred to as fluence) is generally expressed as the product of UV light intensity ( $I$ ) and irradiation time ( $T_{\text{irr}}$ ), according to **Eq. 1**:

$$\text{UV dose} = I \times T_{\text{irr}} \quad (\text{Eq. 1})$$

where UV dose is commonly expressed as  $\text{J/cm}^2 = \text{W sec/cm}^2$ .

It is worthy of note that UV dose is a key parameter when UV radiation (whether or not used in combination with TiO<sub>2</sub>) is used as an antimicrobial mean.

## **2. Antibacterial effects of UV light in combination or not with TiO<sub>2</sub>**

The antimicrobial activity of UV-activated TiO<sub>2</sub> was first demonstrated by Matsunaga and coworkers in 1985 [35]. Since then, a substantial body of literature has addressed the antimicrobial effects of photocatalytic TiO<sub>2</sub> nanoparticles against both Gram negative and Gram positive bacteria. The most relevant examples of TiO<sub>2</sub> nanoparticles and microparticles used in the form of powder, and typically dispersed in aqueous solutions (**Table 1**), or immobilized onto surfaces (**Table 2**) are reported in the tables herein below.



**Table 1.** Studies on the photocatalytic activity of TiO<sub>2</sub> used as nanoparticle and microparticle suspensions against bacteria.

target	[photocatalyst]	light parameters	irradiation time	antibacterial efficiency	Estimated minimum UV dose (according to Eq. 1)	Ref.
<i>S. choleraesuis</i> , <i>V. parahaemolyticus</i> , <i>L. monocytogenes</i>	10 mg/mL (Petri dish)	$\lambda = 360$ nm (UV-A); $I = 0.4$ mW/cm <sup>2</sup>	30 min 1 hr 1.5 hrs 2 hrs	100% at $T_{irr} \geq 2$ hrs	0.3 J/cm <sup>2</sup> (or W sec/cm <sup>2</sup> )	[36]
	0.25 - 1.25 mg/mL (batch reactor)	$\lambda = 360$ nm (UV-A); $I = 0.1$ mW/cm <sup>2</sup>	3 hrs 4 hrs	100% at $T_{irr} \geq 3$ hrs	0.1 J/cm <sup>2</sup>	
<i>E. coli</i>	0.025 - 1 mg/mL	$\lambda = 400 - 800$ nm (Visible – IR); $I = 0.04$ mW/cm <sup>2</sup> $I = 0.1$ mW/cm <sup>2</sup>	2 hrs	100% at $T_{irr} \geq 40$ min 100% at $T_{irr} \geq 25$ min	96 J/cm <sup>2</sup> 150 J/cm <sup>2</sup>	[37]
<i>S. aureus</i> , <i>S. typhimurium</i> , <i>P. aeruginosa</i> , <i>E. coli</i>	1 mg/mL	$\lambda = 368$ nm (UV-A); $I = n.d.$	30 min	100% at $T_{irr} \geq 1$ hr	/	[38]
			1 hr			
			1.5 hrs			
			2 hrs			
			2.5 hrs			



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<i>E. coli</i>	1 mg/mL	$\lambda = 310 - 400 \text{ nm (UV-A)}$ ; $I = 0.5 \text{ mW/cm}^2$	30 min	100% at $T_{irr} \geq 4 \text{ hrs}$	1.8 J/cm <sup>2</sup>	[39]						
			1 hr									
			1.5 hrs									
			2 hrs									
			2.5 hrs									
			3 hrs									
			3.5 hrs									
			4 hrs									
			<hr/>									



**Table 2.** Studies on the photocatalytic activity of TiO<sub>2</sub>-based photoreactors against bacteria.

target	photoreactor	light parameters	irradiation time	antibacterial efficiency	Estimated minimum UV dose (according to Eq. 1)	Ref.
<i>E. coli</i>	TiO <sub>2</sub> -coated Petri dish	$\lambda = 310 - 400 \text{ nm (UV-A)}$ ; $I = 0.25 \text{ mW/cm}^2$	2 hrs 4 hrs 6 hrs	100% at $T_{\text{irr}} \geq 2 \text{ hrs}$	7.2 J/cm <sup>2</sup>	[39]
<i>M. smegmatis</i> , <i>B. thuringiensis</i>	TiO <sub>2</sub> and Pt/TiO <sub>2</sub> -coated glass	$\lambda = 350 - 400 \text{ nm (UV-A)}$ ; $I = 0.65 \text{ mW/cm}^2$	10 min 20 min 30 min	99.8% at $T_{\text{irr}} \geq 30 \text{ min}$	1.1 J/cm <sup>2</sup>	[40]
<i>E. coli</i>	TiO <sub>2</sub> -coated glass	$\lambda = 315-400 \text{ nm (UV-A)}$ ; $I = 1 \text{ mW/cm}^2$	15 min 30 min 1 hr 1.5 hrs	100% at $T_{\text{irr}} \geq 30 \text{ min}$	1.8 J/cm <sup>2</sup>	[41]
<i>E. coli</i>	TiO <sub>2</sub> -coated filter	$\lambda = 355-375 \text{ nm (UV-A)}$ $\lambda = 280-320 \text{ nm (UV-B)}$ $\lambda = 254 \text{ nm (UV-C)}$ ; $I = 3.6 \text{ mW/cm}^2$	2 hrs 4 hrs 6 hrs	100% at $T_{\text{irr}} \geq 4 \text{ hrs}$	518 J/cm <sup>2</sup>	[21]
<i>E. coli</i> , <i>P. aeruginosa</i> , <i>C. freundii</i> , <i>S. aureus</i> , <i>S. saprophyticus</i> MRSA	TiO <sub>2</sub> -coated cellulose acetate monoliths	$\lambda = 365 \text{ nm (UV-A)}$ ; $I = \text{n.d.}$	5 min 10 min 15 min 20 min	100% at $T_{\text{irr}} = 20 \text{ min}$	/	[22]

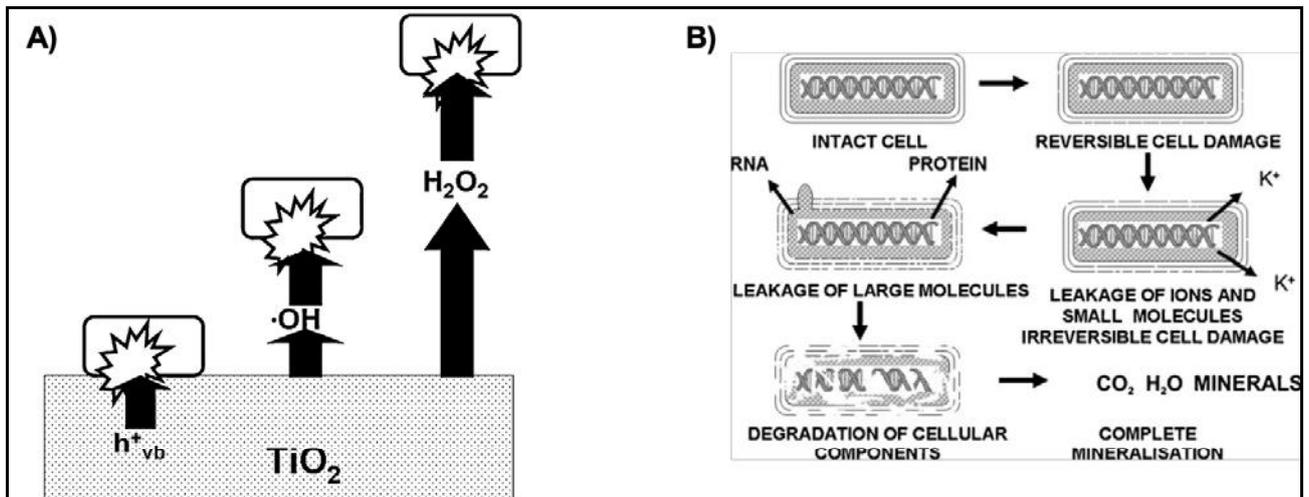


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<i>E. coli</i>	TiO <sub>2</sub> film	$\lambda = 365 \text{ nm (UV-A)}$ ; $I = \text{n.d.}$	1 hr 3 hrs 6 hrs 8 hrs	100% at $T_{\text{irr}} \geq 6 \text{ hrs}$	/	[42]
<i>E. coli</i>	continuous annular reactor with TiO <sub>2</sub> -coated filter	$\lambda = 365 \text{ nm (UV-A)}$ ; $I = 0.5 \text{ mW/cm}^2$ $I = 3.4 \text{ mW/cm}^2$	1.1 min	100%	0.03 J/cm <sup>2</sup> 0.204 J/cm <sup>2</sup>	[24]



Generally speaking, the UV photocatalytic inactivation of bacteria is mainly due to damage of the cell wall, membrane, enzymes, and nucleic acids by ROS and their stable byproducts [43]. In particular, Sunada *et al.* proposed a three-step mechanism for photokilling of bacteria on irradiated TiO<sub>2</sub>-surfaces (**Figure 3**): i) attack of cell walls by ROS; ii) disordering of the inner cytoplasmic membrane and killing of the cell; iii) degradation of the intracellular components [44].



**Figure 3. A)** Role of ROS in the photocatalytic-induced killing mechanism of bacteria. Direct oxidation of cell components can occur when cells are in direct contact with the catalyst. Hydroxyl radicals and H<sub>2</sub>O<sub>2</sub> are involved close to and distant from the catalyst, respectively. **B)** Scheme for photocatalytic killing and destruction of bacteria on TiO<sub>2</sub>. Contact between the cells and TiO<sub>2</sub> may affect membrane permeability (reversible process). This is followed by increased damage to all cell wall layers, allowing leakage of small molecules such as ions. Damage at this stage may be irreversible, and it accompanies cell death. Furthermore, membrane damage allows leakage of higher molecular weight components such as proteins, which may be followed by protrusion of the cytoplasmic membrane into the surrounding medium through degraded areas of the peptidoglycan and cell lysis. Degradation of the internal components of the cell then occurs, followed by complete mineralization. The degradation process may occur progressively from the side of the cell in contact with the catalyst (adapted by [45]).

By contrast, it has been shown that far-UV-C light ( $\lambda = 207 - 222 \text{ nm}$ ; UV dose = 135 mJ/cm<sup>2</sup>) efficiently inactivates drug-resistant bacteria, without apparent harm to exposed mammalian skin [46,47]. Short-wave UV-C radiation is highly disinfectant because the light can efficiently pass through and inactivate microorganisms as their size is typically of  $\mu\text{m}$  or smaller. This means that UV-C has an intensive biocidal effect and may render harmless viruses, bacteria, yeasts and fungi within seconds. Moreover, the microorganisms cannot gain resistance to UV radiation. Based on literature data, the radiant exposure of UV-C (i.e., UV dose) needed for complete sterilization was usually in the order of tens to hundreds of mJ/cm<sup>2</sup> [11,48,49].



### **3. Antiviral effects of UV light in combination or not with TiO<sub>2</sub>**

UV-activated photocatalysis has been found to inactivate mammalian viruses including poliovirus 1, avian and human influenza viruses, and SARS coronavirus, as reported in **Table 3**.

Experimental findings demonstrate that the photocatalysis induced by TiO<sub>2</sub> significantly inactivates the influenza virus by degrading viral proteins, and the degradation depends on the UV-A intensity ( $I$ ) and irradiation time ( $T_{irr}$ ).

Besides, simple exposition to UV light has been reported to be an efficient way to inactivate viruses. It has been shown that effective inactivation occurs under an environmental level of UV-A intensity, as reported in literature [50]. Moreover, using a conventional UV-C lamp at  $\lambda = 254$  nm capable of providing a UV dose of  $1.1 \text{ mJ/cm}^2$ , McDevitt *et al.* [51] found an inactivation of  $\sim 95\%$  of airborne influenza virus H1N1 virus. Similar results were found by Welch D *et al.* [52], but in different conditions (i.e., 30 min-exposure at far UV-C light ( $\lambda = 207 - 222$  nm), corresponding to a very low dose of  $2 \text{ mJ/cm}^2$ ), and by Tsunetsugu-Yokota [53] (i.e., 20 min-exposition at UV-A light intensity of  $1.3 \text{ mW/cm}^2$ , corresponding to a UV dose of  $1.6 \text{ J/cm}^2$ ).



**Table 3.** Studies on the photocatalytic activity of TiO<sub>2</sub>-based photoreactors against viruses.

target	photoreactor	light parameters	irradiation time	antiviral efficacy	Estimated minimum UV dose (according to Eq. 1)	Ref.
influenza virus H1N1	TiO <sub>2</sub> -coated porous ceramic substrate	$\lambda = 365 \text{ nm (UV-A)}$ ; $I = 1 \text{ mW/cm}^2$	5 min	100% at $T_{irr} \geq 5 \text{ min}$	0.3 J/cm <sup>2</sup>	[19]
			10 min			
			15 min			
			30 min			
vaccinia virus, influenza virus H3N3	TiO <sub>2</sub> and Pt/TiO <sub>2</sub> -coated glass	$\lambda = 350\text{-}400 \text{ nm (UV-A)}$ ; $I = 0.65 \text{ mW/cm}^2$	10 min	99.8% at $T_{irr} \geq 30 \text{ min}$	1.1 J/cm <sup>2</sup>	[40]
			20 min			
			30 min			
influenza virus H1N1	TiO <sub>2</sub> -coated glass	$\lambda = 352 \text{ nm (UV-A)}$ ; $I = 0.001 \text{ mW/cm}^2$ $I = 0.01 \text{ mW/cm}^2$ $I = 0.1 \text{ mW/cm}^2$ $I = 1 \text{ mW/cm}^2$	2 hrs	100%	0.8-14.4 J/cm <sup>2</sup>	[50]
			4 hrs			
			6 hrs			
			6 hrs			
			8 hrs			
HSV-1 virus	TiO <sub>2</sub> film	$\lambda = 365 \text{ nm (UV-A)}$ ; $I = \text{n.d.}$	6 hrs	100%	/	[42]
noravirus	TiO <sub>2</sub> photocatalytic reactor	$\lambda = 254 \text{ nm (UV-B)}$	5 min	100% at $T_{irr} \geq 10 \text{ min}$	2.7 J/cm <sup>2</sup>	[54]
			10 min			
			15 min			
			15 min			
			20 min			



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#### 4. Conclusions

Photocatalytic inactivation of microorganisms is a synergistic bactericidal effect of light and oxidative radicals generated by  $\text{TiO}_2$ . The extent of each one varies as a function of different physical parameters.

A positive effect was observed and depended on:

- UV light intensity (optimal range: 0.5 – 3.5  $\text{mW}/\text{cm}^2$ )
- irradiation time (optimal range: 30 min - 2 hrs)
- $\text{TiO}_2$  concentration in solution (optimal range: 0.5 – 1.0  $\text{mg}/\text{mL}$ )

Other parameters which affect the photocatalytic bacterial inactivation are:

- Immobilization of catalyst onto a surface
- The crystalline form of catalyst was studied using suspended and immobilized  $\text{TiO}_2$ .

In conclusion, according to the data shown in the tables herein above, UV-A-induced photocatalysis of  $\text{TiO}_2$  or  $\text{TiO}_2$ -based surface at a suitable light intensity ( $I$ ) has the potential to provide a powerful tool in the fight against transmission of infectious disease in a very short irradiation time ( $T_{\text{irr}}$ ) of about 30 min. As a valuable alternative, inactivation of airborne microorganisms can be reached by using 30-min UV light irradiation time ( $T_{\text{irr}}$ ) across the UV-C spectrum, depending on the UV light intensity ( $I$ ).

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## References

1. Kim, J.; Jang, J. Inactivation of airborne viruses using vacuum ultraviolet photocatalysis for a flow-through indoor air purifier with short irradiation time. *Aerosol Sci. Technol.* **2018**, *52*, 557–566.
2. Xu, Z.; Wu, Y.; Shen, F.; Chen, Q.; Tan, M.; Yao, M. Bioaerosol Science, Technology, and Engineering: Past, Present, and Future. *Aerosol Sci. Technol.* **2011**, *45*, 1337–1349.
3. Candiani, G.; Del Curto, B.; Cigada, A. Improving indoor air quality by using the new generation of corrugated cardboard-based filters. *J. Appl. Biomater. Funct. Mater.* **2012**, *10*, 157–162.
4. Yu, B.F.; Hu, Z.B.; Liu, M.; Yang, H.L.; Kong, Q.X.; Liu, Y.H. Review of research on air-conditioning systems and indoor air quality control for human health. *Int. J. Refrig.* **2009**, *32*, 3–20.
5. Ryan, K.; McCabe, K.; Clements, N.; Hernandez, M.; Miller, S.L. Inactivation of Airborne Microorganisms Using Novel Ultraviolet Radiation Sources in Reflective Flow-Through Control Devices. *Aerosol Sci. Technol.* **2010**, *44*, 541–550.
6. Lee, B.U. Life Comes from the Air: A Short Review on Bioaerosol Control. *Aerosol Air Qual. Res.* **2011**, *11*, 921–927.
7. Hwang, G.B.; Jung, J.H.; Jeong, T.G.; Lee, B.U. Effect of hybrid UV-thermal energy stimuli on inactivation of *S. epidermidis* and *B. subtilis* bacterial bioaerosols. *Sci. Total Environ.* **2010**, *408*, 5903–5909.
8. Hollaender, A.; Du Buy, H.G.; Ingraham, H.S.; Wheeler, S.M. Control of air-borne microorganisms by ultraviolet floor irradiation. *Science (80- )*. **1944**, *99*, 130–131.
9. Kowalski, W. *Ultraviolet germicidal irradiation handbook : uvgi for air and surface disinfection.*; Springer, 2014; ISBN 9783642424809.
10. Kujundzic, E.; Matalkah, F.; Howard, C.J.; Hernandez, M.; Miller, S.L. UV air cleaners and upper-room air ultraviolet germicidal irradiation for controlling airborne bacteria and fungal spores. *J. Occup. Environ. Hyg.* **2006**, *3*, 536–546.
11. Conner-Kerr, T.A.; Sullivan, P.K.; Gaillard, J.; Franklin, M.E.; Jones, R.M. The effects of ultraviolet radiation on antibiotic-resistant bacteria in vitro. *Ostomy. Wound. Manage.* **1998**, *44*, 50–56.
12. Budowsky, E.I.; Bresler, S.E.; Friedman, E.A.; Zheleznova, N. V. Principles of selective inactivation of viral genome - I. UV-induced inactivation of influenza virus. *Arch. Virol.* **1981**, *68*, 239–247.
13. Setlow, R.B.; Grist, E.; Thompson, K.; Woodhead, A.D. Wavelengths effective in induction of malignant melanoma. *Proc. Natl. Acad. Sci. U. S. A.* **1993**, *90*, 6666–6670.
14. Benabbou, A.K.; Derriche, Z.; Felix, C.; Lejeune, P.; Guillard, C. Photocatalytic inactivation of *Escherichia coli*. Effect of concentration of TiO<sub>2</sub> and microorganism, nature, and intensity of UV irradiation. *Appl. Catal. B Environ.* **2007**, *76*, 257–263.
15. Gaya, U.I.; Abdullah, A.H. Heterogeneous photocatalytic degradation of organic contaminants over titanium dioxide: A review of fundamentals, progress and problems. *J. Photochem. Photobiol. C Photochem. Rev.* **2008**, *9*, 1–12.
16. Pelaez, M.; Nolan, N.T.; Pillai, S.C.; Seery, M.K.; Falaras, P.; Kontos, A.G.; Dunlop, P.S.M.; Hamilton, J.W.J.; Byrne, J.A.; O'Shea, K.; et al. A review on the visible light active titanium dioxide photocatalysts for environmental applications. *Appl. Catal. B Environ.* **2012**, *125*, 331–349.
17. Friedmann, D.; Mendive, C.; Bahnemann, D. TiO<sub>2</sub> for water treatment: Parameters affecting the kinetics and mechanisms of photocatalysis. *Appl. Catal. B Environ.* **2010**, *99*, 398–406.
18. Visai, L.; de Nardo, L.; Punta, C.; Melone, L.; Cigada, A.; Imbriani, M.; Arciola, C.R.



- Titanium oxide antibacterial surfaces in biomedical devices. *Int. J. Artif. Organs* **2011**, *34*, 929–946.
19. Daikoku, T.; Takemoto, M.; Yoshida, Y.; Okuda, T.; Takahashi, Y.; Ota, K.; Tokuoka, F.; Kawaguchi, A.T.; Shiraki, K. Decomposition of Organic Chemicals in the Air and Inactivation of Aerosol-Associated Influenza Infectivity by Photocatalysis. *Aerosol Air Qual. Res.* **2015**, *15*, 1469–1484.
  20. Josset, S.; Taranto, J.; Keller, N.; Keller, V.; Lett, M.C. Photocatalytic Treatment of Bioaerosols: Impact of the Reactor Design. *Environ. Sci. Technol.* **2010**, *44*, 2605–2611.
  21. Pigeot-Remy, S.; Lazzaroni, J.C.; Simonet, F.; Petinga, P.; Vallet, C.; Petit, P.; Vialle, P.J.; Guillard, C. Survival of bioaerosols in HVAC system photocatalytic filters. *Appl. Catal. B Environ.* **2014**, *144*, 654–664.
  22. Rodrigues-Silva, C.; Miranda, S.M.; Lopes, F.V.S.; Silva, M.; Dezotti, M.; Silva, A.M.T.; Faria, J.L.; Boaventura, R.A.R.; Vilar, V.J.P.; Pinto, E. Bacteria and fungi inactivation by photocatalysis under UVA irradiation: liquid and gas phase. *Environ. Sci. Pollut. Res.* **2017**, *24*, 6372–6381.
  23. Lin, W.C.; Chen, C.N.; Tseng, T.T.; Wei, M.H.; Hsieh, J.H.; Tseng, W.J. Micellar layer-by-layer synthesis of TiO<sub>2</sub>/Ag hybrid particles for bactericidal and photocatalytic activities. *J. Eur. Ceram. Soc.* **2010**, *30*, 2849–2857.
  24. Pal, A.; Pehkonen, S.O.; Yu, L.E.; Ray, M.B. Photocatalytic inactivation of airborne bacteria in a continuous-flow reactor. *Ind. Eng. Chem. Res.* **2008**, *47*, 7580–7585.
  25. Keller, N.; Rebmann, G.; Barraud, E.; Zahraa, O.; Keller, V. Macroscopic carbon nanofibers for use as photocatalyst support. In *Proceedings of the Catalysis Today*; Elsevier, 2005; Vol. 101, pp. 323–329.
  26. Miyagi, T.; Kamei, M.; Mitsunashi, T.; Ishigaki, T.; Yamazaki, A. Charge separation at the rutile/anatase interface: A dominant factor of photocatalytic activity. *Chem. Phys. Lett.* **2004**, *390*, 399–402.
  27. Fujishima, A.; Honda, K. Electrochemical photolysis of water at a semiconductor electrode. *Nature* **1972**, *238*, 37–38.
  28. Fujishima, A.; Zhang, X. Titanium dioxide photocatalysis: present situation and future approaches. *Comptes Rendus Chim.* **2006**, *9*, 750–760.
  29. Kikuchi, Y.; Sunada, K.; Iyoda, T.; Hashimoto, K.; Fujishima, A. Photocatalytic bactericidal effect of TiO<sub>2</sub> thin films: Dynamic view of the active oxygen species responsible for the effect. *J. Photochem. Photobiol. A Chem.* **1997**, *106*, 51–56.
  30. Sunada, K.; Kikuchi, Y.; Hashimoto, K.; Fujishima, A. Bactericidal and detoxification effects of TiO<sub>2</sub> thin film photocatalysts. *Environ. Sci. Technol.* **1998**, *32*, 726–728.
  31. Kühn, K.P.; Chaberny, I.F.; Massholder, K.; Stickler, M.; Benz, V.W.; Sonntag, H.G.; Erdinger, L. Disinfection of surfaces by photocatalytic oxidation with titanium dioxide and UVA light. *Chemosphere* **2003**, *53*, 71–77.
  32. Yu, J.C.; Ho, W.; Lin, J.; Yip, H.; Wong, P.K. Photocatalytic activity, antibacterial effect, and photoinduced hydrophilicity of TiO<sub>2</sub> films coated on a stainless steel substrate. *Environ. Sci. Technol.* **2003**, *37*, 2296–2301.
  33. Brook, L.A.; Evans, P.; Foster, H.A.; Pemble, M.E.; Steele, A.; Sheel, D.W.; Yates, H.M. Highly bioactive silver and silver/titania composite films grown by chemical vapour deposition. *J. Photochem. Photobiol. A Chem.* **2007**, *187*, 53–63.
  34. Ditta, I.B.; Steele, A.; Liptrot, C.; Tobin, J.; Tyler, H.; Yates, H.M.; Sheel, D.W.; Foster, H.A. Photocatalytic antimicrobial activity of thin surface films of TiO<sub>2</sub>, CuO and TiO<sub>2</sub>/CuO dual layers on *Escherichia coli* and bacteriophage T4. *Appl. Microbiol. Biotechnol.* **2008**, *79*, 127–133.
  35. Matsunaga, T.; Tomoda, R.; Nakajima, T.; Wake, H. Photoelectrochemical sterilization of microbial cells by semiconductor powders. *FEMS Microbiol. Lett.* **1985**, *29*, 211–214.



36. Kim, B.; Kim, D.; Cho, D.; Cho, S. Bactericidal effect of TiO<sub>2</sub> photocatalyst on selected food-borne pathogenic bacteria. *Chemosphere* **2003**, *52*, 277–281.
37. Rincón, A.G.; Pulgarin, C. Photocatalytic inactivation of E. coli: Effect of (continuous-intermittent) light intensity and of (suspended-fixed) TiO<sub>2</sub> concentration. *Appl. Catal. B Environ.* **2003**, *44*, 263–284.
38. Cushnie, T.P.T.; Robertson, P.K.J.; Officer, S.; Pollard, P.M.; McCullagh, C.; Robertson, J.M.C. Variables to be considered when assessing the photocatalytic destruction of bacterial pathogens. *Chemosphere* **2009**, *74*, 1374–1378.
39. Verdier, T.; Coutand, M.; Bertron, A.; Roques, C. Antibacterial activity of TiO<sub>2</sub> photocatalyst alone or in coatings on E. coli: The influence of methodological aspects. *Coatings* **2014**, *4*, 670–686.
40. Kozlova, E.A.; Safatov, A.S.; Kiselev, S.A.; Marchenko, V.Y.; Sergeev, A.A.; Skarnovich, M.O.; Emelyanova, E.K.; Smetannikova, M.A.; Buryak, G.A.; Vorontsov, A. V. Inactivation and mineralization of aerosol deposited model pathogenic microorganisms over TiO<sub>2</sub> and Pt/TiO<sub>2</sub>. *Environ. Sci. Technol.* **2010**, *44*, 5121–5126.
41. Kim, D.; Kwak, S.Y. Photocatalytic inactivation of e coli with a mesoporous TiO<sub>2</sub> coated film using the film adhesion method. *Environ. Sci. Technol.* **2009**, *43*, 148–151.
42. Hajkova, P.; Spatenka, P.; Horsky, J.; Horska, I.; Kolouch, A. Photocatalytic effect of TiO<sub>2</sub> films on viruses and bacteria. *Plasma Process. Polym.* **2007**, *4*, 397–401.
43. Guo, Q.; Xu, C.; Ren, Z.; Yang, W.; Ma, Z.; Dai, D.; Fan, H.; Minton, T.K.; Yang, X. Stepwise photocatalytic dissociation of methanol and water on TiO<sub>2</sub>(110). *J. Am. Chem. Soc.* **2012**, *134*, 13366–13373.
44. Sunada, K.; Watanabe, T.; Hashimoto, K. Studies on photokilling of bacteria on TiO<sub>2</sub> thin film. *J. Photochem. Photobiol. A Chem.* **2003**, *156*, 227–233.
45. Foster, H.A.; Ditta, I.B.; Varghese, S.; Steele, A. Photocatalytic disinfection using titanium dioxide: Spectrum and mechanism of antimicrobial activity. *Appl. Microbiol. Biotechnol.* **2011**, *90*, 1847–1868.
46. Buonanno, M.; Randers-Pehrson, G.; Bigelow, A.W.; Trivedi, S.; Lowy, F.D.; Spotnitz, H.M.; Hammer, S.M.; Brenner, D.J. 207-nm UV Light - A Promising Tool for Safe Low-Cost Reduction of Surgical Site Infections. I: In Vitro Studies. *PLoS One* **2013**, *8*, 1–7.
47. Buonanno, M.; Stanislauskas, M.; Ponnaiya, B.; Bigelow, A.W.; Randers-Pehrson, G.; Xu, Y.; Shuryak, I.; Smilenov, L.; Owens, D.M.; Brenner, D.J. 207-nm UV light - A promising tool for safe low-cost reduction of surgical site infections. II: In-vivo safety studies. *PLoS One* **2016**, *11*, 1–12.
48. Sullivan, P.K.; Conner-Kerr, T.A. A comparative study of the effects of UVC irradiation on select prokaryotic and eukaryotic wound pathogens. *Ostomy. Wound. Manage.* **2000**, *46*, 28–34.
49. Mohr, H.; Steil, L.; Gravemann, U.; Thiele, T.; Hammer, E.; Greinacher, A.; Müller, T.H.; Völker, U. A novel approach to pathogen reduction in platelet concentrates using short-wave ultraviolet light. *Transfusion* **2009**, *49*, 2612–2624.
50. Nakano, R.; Ishiguro, H.; Yao, Y.; Kajioka, J.; Fujishima, A.; Sunada, K.; Minoshima, M.; Hashimoto, K.; Kubota, Y. Photocatalytic inactivation of influenza virus by titanium dioxide thin film. *Photochem. Photobiol. Sci.* **2012**, *11*, 1293–1298.
51. McDevitt, J.J.; Rudnick, S.N.; Radonovich, L.J. Aerosol susceptibility of influenza virus to UV-C light. *Appl. Environ. Microbiol.* **2012**, *78*, 1666–1669.
52. Welch, D.; Buonanno, M.; Grilj, V.; Shuryak, I.; Crickmore, C.; Bigelow, A.W.; Randers-Pehrson, G.; Johnson, G.W.; Brenner, D.J. Far-UVC light: A new tool to control the spread of airborne-mediated microbial diseases. *Sci. Rep.* **2018**, *8*, 1–7.
53. Tsunetsugu-Yokota, Y. Large-scale preparation of UV-inactivated SARS coronavirus virions for vaccine antigen. *Methods Mol. Biol.* **2008**, *454*, 119–126.



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54. Kim, S.H.; Shahbaz, H.M.; Park, D.; Chun, S.; Lee, W.; Oh, J.W.; Lee, D.U.; Park, J. A combined treatment of UV-assisted TiO<sub>2</sub> photocatalysis and high hydrostatic pressure to inactivate internalized murine norovirus. *Innov. Food Sci. Emerg. Technol.* **2017**, *39*, 188–196.